

Pseudo-Prolines: Reversible Conformational Trap of Cyclosporin C as Novel Concept for Prodrug Design

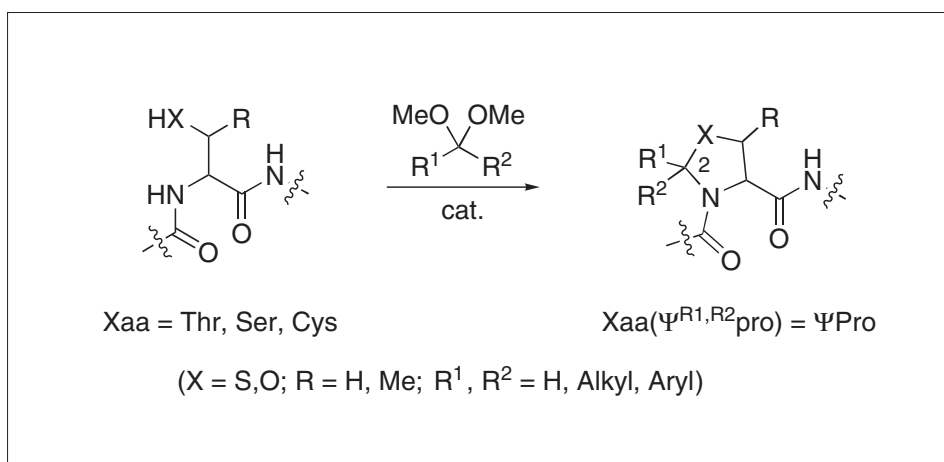
Olivier Turpin*, Manfred Mutter, and Luc Patiny

Abstract: The selective and reversible insertion of pseudo-proline (Ψ Pro) systems in cyclosporin C (CsC) featuring different C(2) substituents at the oxazolidine ring and its impact on the conformational and biological properties is described. The presence of a 5-membered ring exerts drastic effects upon the backbone conformation of CsC as demonstrated by NMR analysis. For example, the number of conformations, in particular in DMSO-d₆, is strongly reduced and a *cis* 1-2 amide bond is induced when dialkylated at the C(2) position, resulting in a complete loss of the binding capacity to its receptor CypA. The reversibility of Ψ Pro insertion allows the temporary introduction of conformational constraints representing a new strategy in pro-drug design.

Keywords: Conformational constraints · Cyclosporin · Pro-drug · Pseudo-proline

Introduction

Our laboratory has developed a new class of proline mimics, referred to as pseudo-prolines (Ψ Pro), for the enhancement of the conformational effects of Pro (Scheme 1). A striking feature of C(2)-dialkylated pseudo-prolines is the induction of a *cis*-amide bond preceding the pseudo-proline unit [1]. As a consequence, the direction of the peptide chain containing such a building block is reversed, resulting in the disruption of secondary structures (notably of β -sheets) and consequently, in the enhancement of solvation effects during peptide synthesis due to the prevention of aggregation caused by hydrophobic interactions [2]. Furthermore, pseudo-prolines were used as a general tool for targeting molecular recognition processes [3]. We have recently demonstrated that Ψ Pro could be directly inserted into complex peptides such



Scheme 1.

as cyclosporin C (CsC). So far we have shown that by reacting CsC with aromatic dimethylacetals, cyclocondensation occurs leading to oxazolidine (Ψ Pro) containing cyclosporin derivatives [4].

Based on these results, we describe in the present article the conformational and configurational aspects resulting from the reversible direct insertion of pseudo-proline (Ψ Pro) [1–3] into cyclosporin C. In particular, the structural impact of a Ψ Pro-induced 1-2 *cis*-amide bond on the overall structure of the cyclic peptide is evaluated by the insertion of various C(2) mono and dialkylated pseudo-proline systems [3a].

Product Synthesis and Characterization

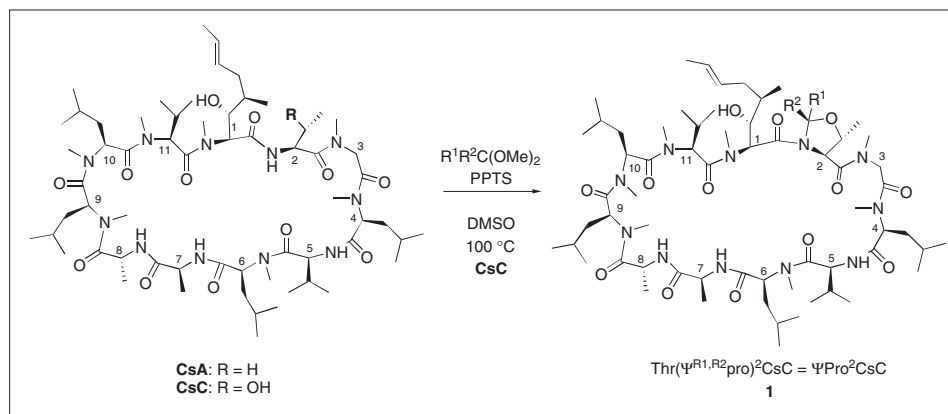
Standard Procedure for the Synthesis of $\text{Thr}(\Psi^{\text{R}^1, \text{R}^2}\text{Pro})^2\text{Cs}$

Dry CsC (100 mg, 0.082 mmol), $\text{R}^1\text{R}^2\text{C}(\text{OMe})_2$ (4.11 mmol, 50 equiv.) and PPTS (6 mg, 0.024 mmol, 0.29 equiv.) in dry DMSO (8.2 ml) was heated to 100–120 °C. After 16 h, the cooled reaction mixture was poured into 500 ml of ethyl acetate/water 1:4 (v/v). The organic layer was washed with brine (5×200 ml) and dried over sodium sulfate. After concentration under reduced pressure, the crude material was pu-

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Table 1. Direct-insertion of Ψ Pro-systems into CsC (see Scheme 2)

| | Compound 1 | | Yield [%] |
|---------------|-----------------|-----------------|-----------|
| | R ¹ | R ² | |
| CsC | – | | |
| 1a | Me | H | 33 |
| 1b | Et | H | 34 |
| 1c | ⁿ Pr | H | 39 |
| 1d | ⁱ Pr | H | 19 |
| 1e | ^t Bu | H | 0 |
| 1f (S) | Ph | H | 61 |
| 1f (R) | H | Ph | 39 |
| 1i | Me | Me | 26 |
| 1j | Et | Et | 22 |
| 1k | ⁿ Pr | ⁿ Pr | 12 |
| 1l | Ph | Ph | 9 |
| 1m | ⁱ Pr | ⁱ Pr | 0 |
| 1n | CF ₃ | CF ₃ | 0 |



Scheme 2.

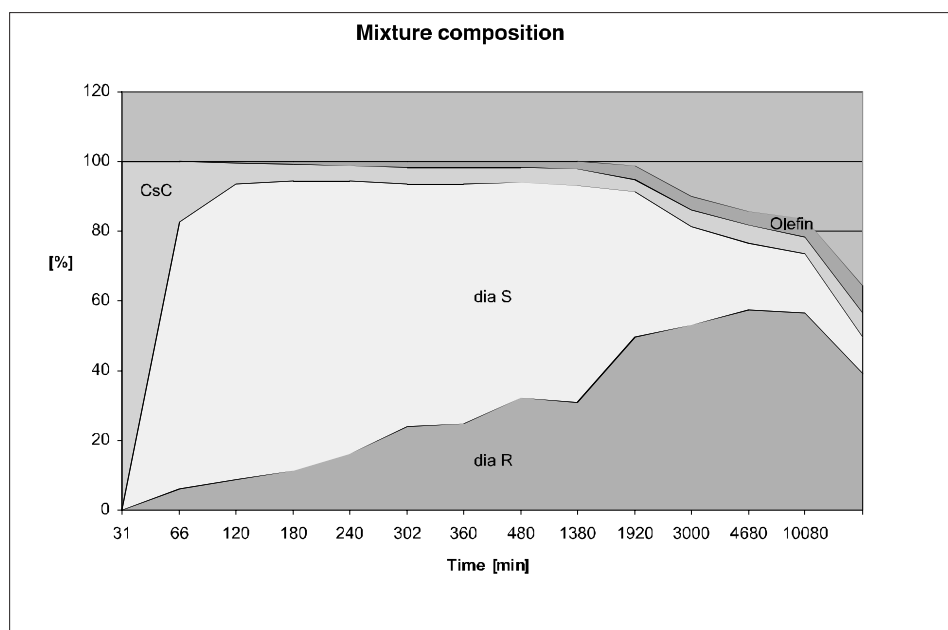


Fig. 1. Time-dependent evolution of the composition of the reaction mixture (**1f**). The kinetically favored (*S*)-diastereoisomer initially obtained is slowly converted to the (*R*)-diastereoisomer (thermodynamic product).

rified by chromatography on silica (acetone/hexane, 4:6) to yield Thr($\Psi^{\text{R}^1, \text{R}^2}\text{Pro}$)²CsC as a white powder.

All compounds were characterized by HPLC, mass spectroscopy, and ¹H NMR.

Results and Discussion

CsC (Scheme 2) differs from the well-known immunosuppressive analogue **CsA** by the presence of a trifunctional amino acid at position 2, *i.e.* a threonine (Thr²) residue replacing 2-amino butyric acid (Abu²) in **CsA**. While the biological and pharmacokinetic properties of **CsC** and **CsA** are very similar [5], the presence of the OH group of Thr² together with a non-methylated amide between residues 1 and 2 renders **CsC** a most attractive candidate for applying the Ψ Pro concept to study conformational changes [3a][4a].

Despite the steric constraints involved in the cyclocondensation of **CsC** with alkyl acetal derivatives (Scheme 2, Table 1, **1a–d**), the direct-insertion proceeds very selectively in one single step with acceptable yields (19–39%). Even strongly hindered oxazolindines derived from dimethylated, diethylated and di-*n*-propylated ketals were obtained in yields ranging from 12 to 26% (**1f–h**). The somewhat lower yields compared to the C(2)monosubstituted derivatives [3a] are indicative of the increased conformational constraints present in the target compounds **1f–i**. Furthermore, the synthesis of extremely sterically demanding oxazolindines such as C(2)-*tert*-butyl, -di(trifluoromethyl) and -diisopropyl derivatives *via* direct insertion of the corresponding substituted ketals proved to be impossible under the established experimental conditions.

In the case of C(2)-monosubstituted pseudo-prolines, alkyl derivatives yield exclusively the (*S*) epimer while in the case of aryl derivatives both epimers are observed. The ratio between the diastereoisomers (*S*) (kinetic derivative) and (*R*) (thermodynamic derivative) could be modulated by changing reaction time and temperature (Fig. 1).

The influence of the insertion of a Ψ Pro on the backbone conformation was investigated using 1D and 2D (TOCSY, ROESY, COSY-DQF, and HSQC) NMR spectroscopy. Because of the pronounced hydrophobicity and insolubility in D₂O of the target molecules, CDCl₃ and DMSO-*d*₆ resembling physiological conditions to some extent [6], were used as solvent. Two categories of products have been investigated. Category one includes the C(2)-monosubstituted Ψ Pro derivatives **1a–h** which show only one conformation in CDCl₃ but multiple conformations in DMSO-*d*₆. Category two includes C(2)-disubstituted Ψ Pro derivatives **1i–l** comprising a limited number

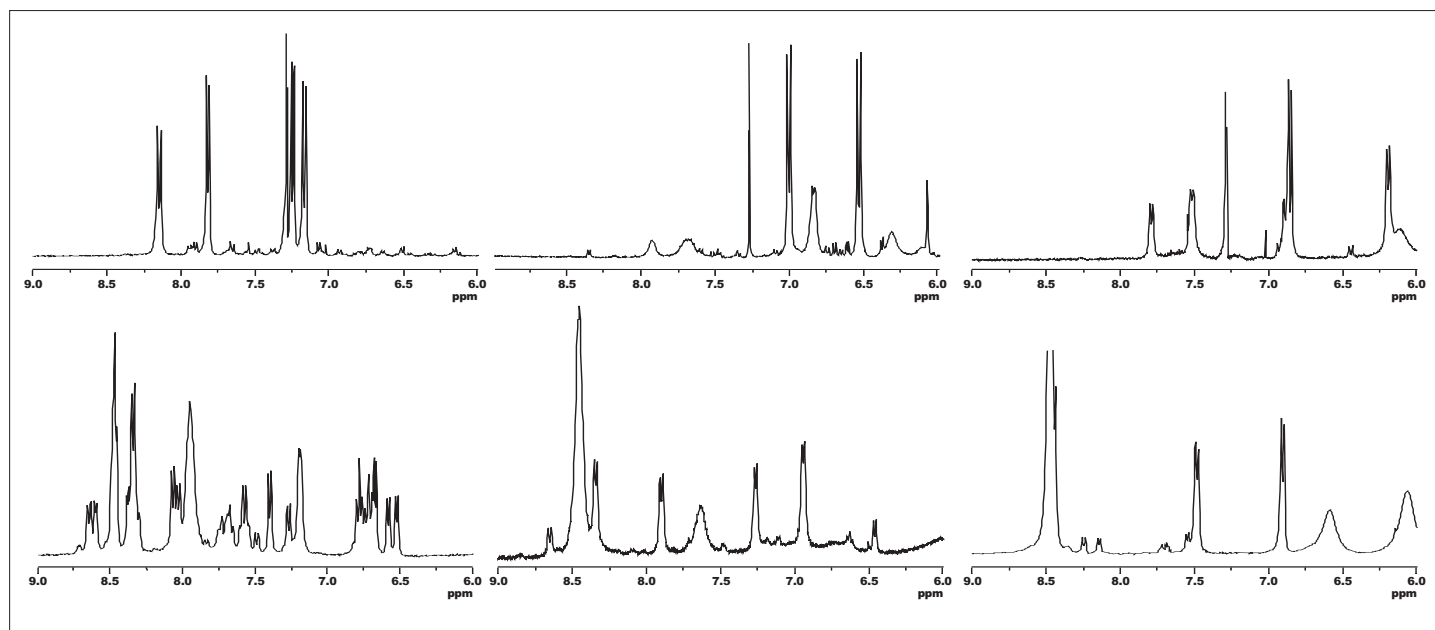


Fig. 2. ¹H NMR of the NH region of CsC (left), **1a** (middle) and **1i** (right) in CDCl₃ (top) and DMSO-d₆ (bottom)

of conformations in CDCl₃ as well as in DMSO-d₆ (Fig. 2). These results point to a striking similarity in the conformational properties of monosubstituted ΨPro-CsA derivatives and CsC. However, the presence of disubstituted ΨPro building blocks rigidifies the cyclosporin ring to such extent that only two or three conformations are accessible even in DMSO-d₆.

The presence of *cis/trans* amide bonds was determined using ROESY experiments. A *cis*-amide bond is characterized by a strong correlation between two consecutive H_α. This is exemplified for derivative **1i** in DMSO-d₆ showing that the two substituents at position C(2) of ΨPro induce a 1-2 *cis*-amide bond in both conformers (Fig. 3), in agreement with previous studies on model dipeptides of the type Xaa-ΨPro [7]. The main difference between the two conformers arises from amide bond 3-4, which was found to be *trans* in the major and *cis* in the minor conformer, whereas the remaining amide bonds exhibit exclusively the *trans* conformation. This result demonstrates that even in cyclic peptides the presence of a dimethylated ΨPro induces a *cis*-amide bond to more than 50%. More bulky alkyl chains at the C(2) position enhance this effect even further as shown for the di-*n*-propyl derivative **1i**.

In the case of C(2)-monosubstituted pseudo-proline derivatives, a new asymmetric center is generated during the reaction. The relative configuration was determined by 2D-NMR spectroscopy based on a ROESY spectrum. Here, a strong NOE effect was observed between 2H_β and 2H_{2α} indicating an (*S*)-C(2) asymmetric carbon (Fig. 4). Interestingly, this observation is

different from the conformational effects of ΨPro-systems in model dipeptides [8] and derivatives obtained by direct-insertion of aromatic ΨPro systems into CsC [3a][4a] where both epimers are present.

Stability of ΨPro

ΨPro-derivatives **1a–n** exhibit differential acid stability, depending on the character of the C(2)-substitution of the oxazoli-

dine system. For the evaluation of the chemical stability of the ΨPro-ring, the compounds were subjected to conditions resembling the native environment. For example, in a mixture of fetal bovine serum/methanol 1:1 (v/v), **1i** proved to be completely stable (>1 month at 37 °C). In 1M HCl_{aq}/acetonitrile 1:1 (v/v) and in 0.1M HCl_{aq}/acetonitrile 1:1 (v/v), half-lives of around 83 min and 17 h, respectively, were determined as monitored by analytical HPLC (Fig. 5). Under the hydrolysis condi-

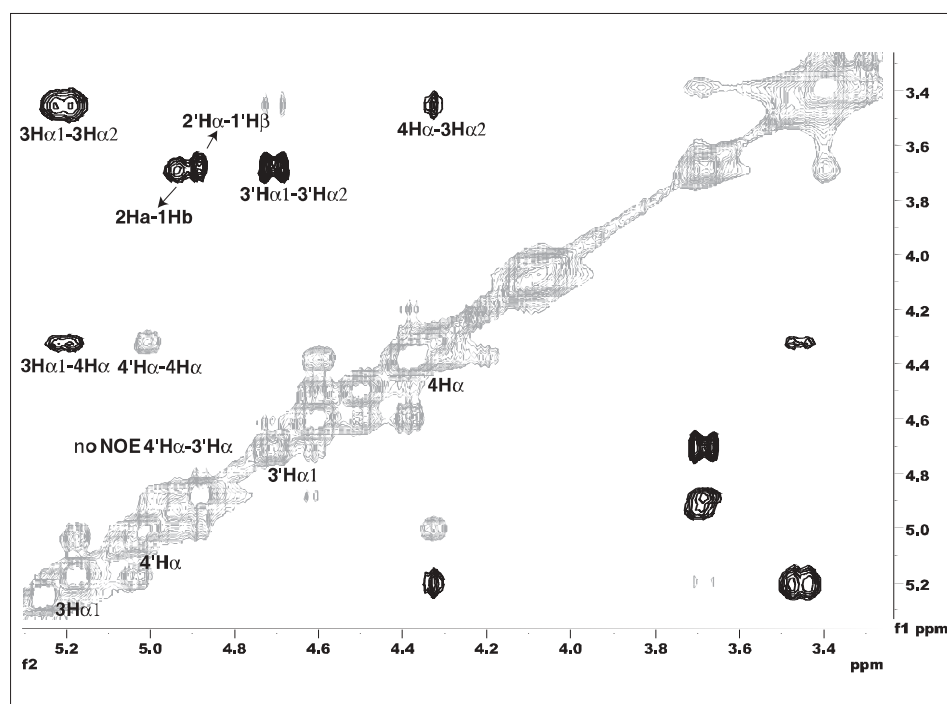


Fig. 3. ROESY of **1i** in DMSO-d₆. The major conformer has 1-2 and 3-4 *cis*-amide bonds (NOE 2H_α-1H_β and 3H_α-4H_α). The minor conformer shows only one *cis*-amide bond 1-2 (NOE 2'H_α-1'H_β, no NOE between 3'H_α and 4'H_α).

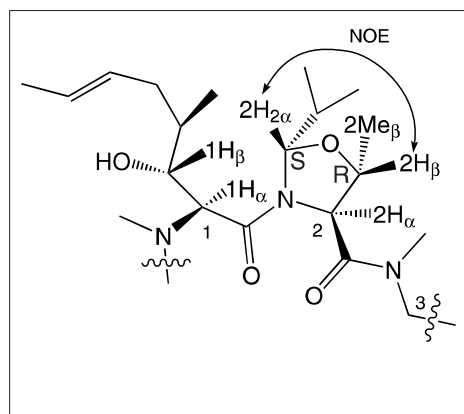


Fig. 4. ROESY experiment for the determination of the relative configuration of C(2) for the ¹³C derivative **1d**.

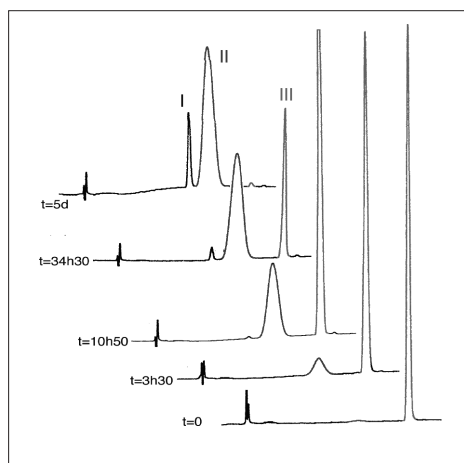


Fig. 5. Hydrolysis of **1i** in 0.1M HCl/acetonitrile 1:1 (v/v) (I: isoCsC, II: CsC, III: **1i**).

tions applied, the restored parent CsC slowly converted to isoCsC as described elsewhere [9].

At pH 6.6, all derivatives except **1g(S)** proved to be stable for several days. The stability of Ψ Pro-containing cyclosporins in water is strongly related to the capacity of the substituents to stabilize the cation at the C(2) position *i.e.* Me (**1a**) Ph (**1f(S)**) (Table 2). Most notably, the presence of a R² substituent increases the stability of the derivatives drastically (Table 2). Shielding of the oxygen and thus hampering protonation as the first step of the hydrolysis cascade might be a plausible explanation. In the case of R² substituted pseudo-prolines, the oxygen is surrounded by neighboring groups on both sides of the ring while in monosubstituted Ψ Pro one face of the oxygen can be freely approached by the proton.

The *in vitro* activity of cyclosporin derivatives was assessed by using the IL-2 reporter gene assay. The immunosuppressive activity of the derivatives is determined as substances interfering with the activation of the T cell signaling cascade inhibit IL-2

Table 2. Hydrolysis kinetics of representative pseudo-proline containing cyclosporins

| | Compound 1 | | Half-life time |
|--------------|-----------------|-----------------|----------------|
| | R ¹ | R ² | pH 1 [h] |
| 1a | Me | H | 7 |
| 1d | ⁱ Pr | H | 9.5 |
| 1f(S) | Ph | H | 1.7 |
| 1f(R) | H | Ph | 49 |
| 1i | Me | Me | 17 |
| 1k | ⁿ Pr | ⁿ Pr | 27 |
| 1l | Ph | Ph | 27.5 |

gene transcription and thus IL-2 production [10]. Furthermore, the binding affinity to CypA was assessed by using the improved spectrophotometric assay described by Rich and coworkers [11]. As expected from the observed conformational constraints induced by the insertion of Ψ Pro systems into CsC, derivatives **1a–n** proved to be trapped into a non-bioactive conformation thus providing a versatile target for prodrug design.

Conclusions

In summary, the incorporation of Ψ Pro systems into CsC results in a pronounced reduction of the conformational space compared to the parent molecule. In tailoring the steric and electronic features of the C(2) substituents, differential acid stability of the Ψ Pro containing CsC derivatives is achieved, paralleled by a complete loss of their receptor binding capacity. Consequently, in applying polar C(2) substituents, the present concept gives access to a new class of water-soluble prodrugs ("soft prodrugs") of considerable therapeutic relevance.

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- [1] a) A. Nefzi, K. Schenk, M. Mutter, *Protein and Peptide Letters* **1994**, *1*, 66; b) T. Wöhr, M. Mutter, *Tetrahedron Lett.* **1995**, *36*, 3847; c) M. Keller, C. Sager, P. Dumy, M. Schutkowski, G.S. Fischer, M. Mutter, *J. Am. Chem. Soc.* **1998**, *120*, 2714.
- [2] a) T. Wöhr, F. Wahl, A. Nefzi, B. Rohwedder, T. Sato, X.C. Sun, M. Mutter, *J. Am. Chem. Soc.* **1996**, *118*, 9218; b) M. Mutter, A. Nefzi, T. Sato, X. Sun, F. Wahl, T. Wöhr, *Peptide Res.* **1995**, *8*, 145.
- [3] a) A. Wittelsberger, M. Keller, L. Scarpellino, L. Patiny, H. Acha-Orbea, M. Mutter, *Angew. Chem. Int. Ed. Engl.* **2000**, *39*, 1111; b) M. Keller, C. Boissard, L. Patiny, N.N. Chung, C. Lemieux, M. Mutter, P.W. Schiller, *J. Med. Chem.* **2001**, *44*, 3896.
- [4] a) M. Keller, T. Wöhr, P. Dumy, L. Patiny, M. Mutter, *Chem. Eur. J.* **2000**, *6*, 4358; b) L. Patiny, J.-F. Guichou, M. Keller, O.

- Turpin, T. Ruckle, P. Lhote, T.M. Buetler, U.T. Ruegg, R.M. Wenger, M. Mutter, *Tetrahedron* **2003**, *59*, 5241.
- [5] a) M. Dreyfuss, E. Harri, H. Hofmann, H. Kobel, W. Pache, H. Tschertter, *European J. Appl. Microbiol.* **1976**, *3*, 125; b) H. Kobel, R. Traber, *European J. Appl. Microbiol. Biotechnol.* **1982**, *14*, 237.
 - [6] a) R. Wenger, J. France, G. Bovermann, L. Walliser, A. Widmer, H. Widmer, *Actual. Chim. Théor.* **1993**, *21*, 95; b) R.M. Wenger, J. France, G. Bovermann, L. Walliser, A. Widmer, H. Widmer, *Febs. Lett.* **1994**, *340*, 255.
 - [7] P. Dumy, M. Keller, D.E. Ryan, B. Rohwedder, T. Wöhr, M. Mutter, *J. Am. Chem. Soc.* **1997**, *119*, 918.
 - [8] a) M. Keller, M. Mutter, C. Lehmann, *Synlett* **1999**, 935; b) M. Keller, C. Lehmann, M. Mutter, *Tetrahedron* **1999**, *55*, 413.
 - [9] R. von Traber, M. Kuhn, A. Rüegger, H. Lichti, H.-R. Loosli, A. von Wartburg, *Helv. Chim. Acta* **1977**, *60*, 1247.
 - [10] a) G. Baumann, G. Zenke, R.M. Wenger, P. Hiestand, V. Quesniaux, E. Andersen, M. Schreier, *J. Autoimmun.* **1992**, *5*, 67; b) P.S. Mattila, K.S. Ullman, S. Fiering, E.A. Emmel, M. McCutcheon, G.R. Crabtree, L.A. Herzenberg, *EMBO J.* **1990**, *9*, 4425.
 - [11] a) J.L. Kofron, P. Kuzmic, V. Kishore, E. Colon-Bonilla, D.H. Rich, *Biochemistry* **1991**, *30*, 6127; b) J.L. Kofron, P. Kuzmic, V. Kishore, G. Gemmecker, S.W. Fesik, D.H. Rich, *J. Am. Chem. Soc.* **1992**, *114*, 2670.